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# REVIEW ON BACTERIOPHAGE THERAPY: RENEWED APPROACH FOR MANAGEMENT OF BACTERIAL INFECTION

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### **Abstract**

Bacteriophages (phages) are viruses that infect and multiply in host bacteria and archaea. They are exceptionally diversified, abundant, and extensive. Phages were discovered just over a century ago and have been used to treat bacterial infections in humans and other animals since the 1919s. This seminar paper examines the usage of bacteriophages over time, their comparative benefits and drawbacks, and their application as a therapeutic tool in animals. Phage therapy is the use of lytic bacteriophages as a treatment against bacterial infections. It was frequently utilized in the pre-antibiotic era until large-scale antibiotic manufacture began. Today, the rising antibiotic resistance dilemma has revitalized bacteriophage therapy.

It has been offered as a possible alternative to antibiotics in the treatment and prevention of human and animal diseases. Its benefits include the fact that it does not harm natural microflora, is relatively safe, and has a positive effect on antibiotic resistance bacteria. Its main disadvantages are a limited host range and the potential of pathogenic gene transmission. Phage therapy in animal production has been debated in veterinary medicine for decades. Several investigations and clinical trials have supported the use of phages as a therapeutic technique for bacterial illnesses in animals. In poultry, swine, cattle, and fish, phage therapy produced good outcomes in terms of lowering mortality, clinical severity, and tissue-level bacterial counts.

It found that phage therapy has a promising outcome for the treatment of bacterial infections in both human and veterinary medicine. However, various obstacles must be resolved before lytic phages may be widely used for therapeutic purposes. To fully benefit from bacteriophage therapy, well-organized research and significant scholarly efforts are required.

Keywords: Antibiotic resistance, Bacteriophages, Pathogenic bacteria, Phage therapy, Virus

### Introduction

Antimicrobial resistance (AMR) is an increasingly serious threat to human and animal population in this scenario. The problem is so serious that treatment of infectious diseases with antibiotics is becoming critically challenging and large number of patients dies annually from untreatable infections (Levy and Marshal, 2004). AMR also has a considerable economic impact: extra hospital costs and associated productivity losses. Looked for huge loss of life and the consequent economic loss and the situation is about to deteriorate even further, as there are only a few drugs left to treat multidrug-resistant bacterial strains led to an interest in discovering new therapeutic tools that allow replacing or complementing antimicrobials when combating bacterial diseases (Magoriakos *et al.*, 2012).

Various modalities have been made to address the problem. These range from the more prudent use of existing antibiotics to the implementation of different antibacterial alternatives such as immune modulators, vaccines, avian egg antibodies, probiotics, and bacteriophage therapy (Brackman et al., 2011). Bacteriophage therapy seems to be promising and sustainable in the long term and implementable in the near future. Indeed, the use of bacteriophages to kill specific bacterial pathogens without harming the majority of the commensal bacteria has received increasing attention during the past decade (Sulakvelidze et al., 2001).

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Bacteriophages are viruses that infect and replicate within host bacteria and archaea (Lyon, 2017). It has relatively simple structures composed of proteins (60%) that encapsulate a DNA or RNA genome (40%). Phages are among the most abundant entities in the biosphere. It has been estimated that there are more than 10^31 population on the planet with a total weight of 10^9 tons. They are ubiquitously distributed in environments populated by bacterial hosts, including soil, water, air, and the intestines of humans and other animals. Phages were usually classified into 19 families, over 140 genera, and more than 5300 types of phage species (Korotyayev and Babichev, 2002). Phages can also be divided into lytic phages and temperate phages based on whether or not their genome is integrated into the bacterial genome (Bao et al., 2018).

Phage therapy is the therapeutic use of phages to treat pathogenic bacterial infections. It was developed and widely used for the treatment of bacterial infection in human medicine, veterinary medicine, and agriculture between 1920 and 1940 until the first large-scale production of the antibiotic (Williamson et al., 2017). Nevertheless, research on phages and their medical applications continued particularly in the former Soviet Union and Eastern Europe. More recently, with the emergence of multiple antibiotic-resistant bacteria, the interest in phage therapy was renewed. Phage therapy has been proposed as a potential candidate to serve as an alternative to antibiotics in the control and prevention of human and animal diseases (Sulakvelidze et al., 2001).

The main advantage of phages is their specificity for target bacteria which reduces the damage to normal flora of the host greatly. Replication at the site of infection is another advantage of phages. They are safe with no or fewer side effects. After their administration phages can dissipate very quickly throughout the body reaching almost every organ and increasing in number whenever host bacteria are present (Clark and Marc, 2006). Bacterial resistance to phages, although likely to arise, should not be a major concern, certainly compared with bacterial resistance to antibiotics. This is because phages grow exponentially, essentially shadowing the bacterial growth and thereby mutating at the same rate due to the plethora of phages, there will certainly be a species that can attack mutated and resistant bacteria to prevent the escape of bacteria and its critical tools to combating antibiotic resistance (Dabrowska et al., 2005)

An increase in bacterial resistance to many antimicrobials has been observed, becoming a subject of global concern in human and veterinary medicine, and led to an interest in discovering new therapeutic tools that allow replacing or complementing antimicrobials when combating bacterial diseases and mitigating antimicrobial resistance crisis. Bacteriophages and viruses that infect and lyse bacteria are among these therapeutic tools. It was widely used before the advent of antibiotics and currently attracting the interest of

the international scientific community (Boerlin, 2010). Therefore, this seminar paper was done with the objective of:

To highlight features of bacteriophages

To provide an insight into bacteriophage therapy, as an alternative to antibiotics and a tool for the mitigation of antibiotic resistance

#### 2. LITERATURE REVIEW

### 2.1. Bacteriophage Definition and Discovery

Bacteriophages (or phages) are viruses that infect and replicate within host bacteria and archaea. The viruses of Kingdom Bacteria were first described as invisible entities capable of destroying bacterial cultures and would remain infectious even after suspensions were passed through filters designed to remove bacteria. Since the action of bacterial viruses resembled the eating of bacterial cultures, the word "phage", which means to eat or devour in Greek, was chosen to describe this phenomenon (Dewangan et al., 2017).

Since ancient times, reports of river waters having the ability to cure infectious diseases have been documented, such as leprosy. In 1896, Ernest Hanbury Hankin reported that something in the waters of the Ganges and Yamuna rivers in India had marked antibacterial action against cholera and could pass through a very fine porcelain filter (Summers, 1999). In 1915, British bacteriologist Frederick Twort discovered a small agent that infected and killed bacteria. He believed the agent must be one of the following: a stage in the life cycle of the bacteria, an enzyme produced by the bacteria themselves, or a virus that grew on and destroyed the bacteria. Independently, French-Canadian microbiologist Félix d'Hérelle announced on 3 September 1917, that he had discovered "an invisible, antagonistic microbe of the dysentery bacillus". D'Hérelle called the virus a bacteriophage. It was D'Hérelle who conducted much research into bacteriophages and introduced the concept of phage therapy (Keen, 2012)

### 2. 2. Ecology and Source of Bacteriophage

Bacteriophages are found in almost all environments on Earth, ranging from soil, sediments, water (both river and seawater), and in/on living or dead plants/animals. Essentially, phages can be isolated from almost any material that will support bacterial growth. The estimated global phage population size is extraordinarily high (Parisien et al., 2008). It has been established that the population number of phages in aquatic systems lies within the range of 104 to 108 virions per ml and about 109 virions /g in the soil, 2 with an estimated total number of 1032 bacteriophages on the planet (Hanlon, 2007).

The production and distribution of phage are dependent on host concentration. The phage populations can grow faster

when there is a greater density of susceptible bacteria. Generally, the virus-to-bacterium ratio falls between 3 and 10, depending on the nutrient level (Brüssow and Kutter, 2005). The consequence of phage lysis not only reduces the productivity of bacterial populations but also delays the ecosystem nutrient cycling and energy flow (Abedon, 2006)

### 2.3. Bacteriophage therapy

### 2.3.1. Definition Bacteriophage Therapy

Phage therapy is the therapeutic use of bacteriophages to treat pathogenic bacterial infections (Brüssow, 2005). Phage therapy may be defined more broadly than just the application of phages to human and animal bodies to combat bacterial disease. Indeed, at its most inclusive phage therapy represents the application of specific phages, which are pathogens of bacteria to selectively reduce or eliminate susceptible bacteria from natural environments, bodies of humans, animals, and artificial environments. In other words, phage therapy is simply another form of biological control the use of one organism to suppress another; and like other biological controls (Sulakvelidze and Morris, 2001)

## 2.3.2. Bacteriophage therapy mode of action and safety profile

Despite the large number of publications on phage therapy, there are very few reports in which the pharmacology of therapeutic phage preparations is delineated. The job of antibacterial, as expressed in terms of pharmacodynamics, is to kill or at least prevent the replication of one or more bacteria while not excessively harming the so-treated patient (Harper, 2006).

Antibiotics possess a wide array of mechanisms of action for example, penicillin inhibits cell wall formation while quinolones inhibit DNA gyrase and topoisomerase in bacteria As for their bactericidal activity, therapeutic phages were assumed to kill their target bacteria by replicating inside and lysing the host cell via a lytic cycle. However, subsequent studies revealed that many therapeutic phages may act via a similar cascade; however, it is also possible that some therapeutic phages have some unique yet unidentified genes or mechanisms responsible for their ability to effectively lyse their target bacteria. For example a group of authors from the EIBMV identified and cloned an anti-Salmonella phage gene responsible, at least in part, for the phage's potent lethal activity against the Salmonella enterica serovar Typhimurium host strains ( Harper, 2006; Leitenberg, 2001)

The most important aspect of phages as antibacterials, other than their ability to kill bacteria, is their low toxicity. This low toxicity is a consequence of phage composition which for tailed phages is almost entirely protein and DNA. Phage interaction with the body's metabolism, such as that associated with the stomach, liver, and various aspects of the animal immune system can readily result in the

degradation of phage virions However, unlike the metabolic decay of certain drugs, phage degradation does not result in the production of toxic byproducts. In addition, intact phages are not highly effective at interacting with other aspects of the body's metabolism such as in terms of specifically binding to or otherwise manipulating body tissues (Merril, 2008).

Furthermore, the body is routinely exposed to large numbers of endogenous phages meaning that the therapeutic application of phages is not qualitatively outside of the normal body experience (Gorski and Weber-Dabrowska, 2005) An exception to this relative safety could be anaphylactic immune-system responses to the proteins associated with phage virions, though even here reactions appear to be rare, plus mild when they do occur (Sulakvelidze and Kutter, 2005). Also, phages can be chosen and/or purified such that they are minimally associated with virulence factors, such as endotoxins or exotoxin-encoding genes (Waldor et al., 2005)

### 2.3.3. Bacteriophage Commercial Product and Legal Status

Phages are not specifically classified as living or chemical agents in any national medicinal legislation. This considerably complicates the regulation of phage therapy clinical trials and commercializing phage products (Fauconnier, 2017). Several clinical trials and case studies known to use bacteriophage therapy have been carried out under different jurisdictions. Individual countries have their regulators, and levels of regulation can be highly variable. (Verbeken et al., 2007)

The European Medicines Agency (EMA) considers bacteriophages biological as agents. Although bacteriophage therapy falls under the existing European regulatory framework on biological medicinal products the directive does not fully cover aspects specific to bacteriophages. Products such as vaccines (some of which are live viruses) that do not have specific regulatory guidelines have been approved by EMA. In the US, similar to the situation in Europe, no bacteriophage therapeutics guidelines for human use have been published by the Food Drug Administration (FDA). Nevertheless, bacteriophage applications are handled by the Division of Vaccines and Related Product Applications (Verbeken et al., 2007; WMA Declaration of Helsinki, 2008)

The FDA currently recognizes commercial bacteriophage preparations against common bacterial pathogens such as Listeria monocytogenes and E. coli as safe and approves their use in food consumed by humans. The first approved food safety-related bacteriophage product was ListShield<sup>TM</sup> (LMP-102<sup>TM</sup>), from Intralytix Inc, a phage cocktail that targets L. monocytogenes contaminants on ready-to-eat (RTE) foods containing meat and poultry products (Bren, 2007). Similar food safety applications and other nonhuman

applications in the agricultural, animal husbandry, veterinary, and diagnostics sectors appear to be progressing well with an increasing number of products becoming available. Such products include AgriPhage, BioTector, EcoShield, Finalise, ListShield, and LISTEXTM P100L (Monk et al., 2010; Ryan et al., 2011).

### 2.4. Advantage and Draw Back Bacteriophage Therapy as Compared to Antibiotic

### 2.4.1. Potential advantage of bacteriophage therapy

Phage therapy, like any other therapeutic method, has advantages and disadvantages mainly related to antibiotic therapy. Among the advantages, the following are described:

- 1. Exclusively bactericidal capacity: bacteria that have been successfully infected by lytic phages are unable to regain viability. In contrast, few antimicrobials have bacteriostatic action only, and as a result, they may allow the evolution of bacterial resistance (LocCarrillo and Abedon, 2011).
- 2. Minimal effects on normal microflora: because of their close host specificity, which may include the ability to infect a few strains or bacterial species, more rarely, the ability to infect more than one genus, closely related to each other. In contrast, many antimicrobial chemicals that possess a wide spectrum of activity are likely to generate superinfections (Loc-Carrillo and Abedon, 2011).
- 3. Reduced potential to induce bacterial resistance: due fact that bacteriophages could have an improved efficacy as compared with antibiotics provides the greatest hope for the future. Regarding this subject, antibiotics have a clear limitation because they are stable, immutable chemicals and therefore are unable to adapt to bacterial mutations (Carlton, 1999)
- 4. Lack of cross-resistance to antibiotics: because phages infect and kill bacteria using different mechanisms to those of antibiotics. Consequently, phages can be effectively used to treat antibiotic-resistant infections such as those caused by multidrug-resistant Staphylococcus aureus (Loc Carrillo and Abedon, 2011).
- 5. Nontoxic effects: some toxicity studies performed with phages in experimental animals such as chickens and mice have shown no toxic effects or adverse reactions in animals (Xie et al., 2005; Gill et al., 2006). Furthermore, they are recognized to be harmless to humans and animals, since recently their use has been approved as an additive in human foods and for direct application in animals (Carlton et al., 2005).
- 6. Possible phage transfer between individuals: this is essentially a cross-infection of phages from treated subjects or environments to untreated individuals, which may be potentially useful in agricultural applications (Rozema et al., 2009).

- 7. Low environmental impact: because of their chemical composition and their narrow host range, phage eliminated after treatment, unlike broad-spectrum antibiotics, in the worst scenario will impact only a small group of environmental bacteria (Loc-Carrillo and Abedon, 2011).
- 8. Low cost: the production of phages predominantly involves growth in its host and further purification. Overall production costs of phages, per unit, do not compare to the costs of pharmaceutical production, while the cost of discovery, isolation, and characterization can be relatively low (Loc-Carrillo and Abedon, 2011).

### 2.4.2. Potential drawback of bacteriophage therapy

Otherwise, the disadvantages of using phages as therapeutic agents the following are described:

- 1. Existence of phage-resistant bacteria: bacteria can evolve resistance to bacteriophages through a variety of mechanisms, including the blocking of viral adsorption, inhibition of the viral genome injection, restriction-modification systems mediated by enzymes that degrade viral nucleic acids or the CRISPR-Cas system (Bikard and Marraffini, 2011), and infection abortion systems of resistance conferred by the Abi system (Buckling and Brockhurst, 2012).
- 2. Not all phages are good therapeutic agents: good therapeutic phages must have a high potential to reach and then kill the bacteria, along with a low potential to modify adversely the environments in which they are applied (Loc-Carrillo and Abedon, 2011).
- 3. Narrow host range: the narrow host range of phages could constitute, at least, a limitation for presumptive treatment. However, as phages may be used in combination with other antimicrobial agents, including other phages, the lytic spectrum of these particles can be much broader than the spectrum of activity of a single phage (Loc-Carrillo and Abedon, 2011).
- 4. Need for high bacterial concentration: this is a needed requirement for the phage to replicate and lyse bacteria. If they are administered in a hurry they will tend to inactivate due to lack of bacteria and higher concentrations will be needed later (Payne and Jansen 2001).
- 5. Interaction of bacteriophages with the innate immune system: may lead to low efficiency of phage therapy (Dombrowsk et al., 2005).
- 6. Existence of physical and chemical barriers that may reduce the phage-bacterium interaction: an example of this has been observed in orally administered phage therapies, where the gastrointestinal environment determines a decrease in the effective encounter between the bacteriophage and the challenge strain (Sklar and Joerger, 2001) since the virus can experience structural problems due to action of digestive enzymes and pH conditions found in the gastrointestinal tract (Higgins et al., 2007).

- 7. Limited knowledge about the pharmacology of phage: this proves to be another big issue (Dabrowska et al., 2005)
- 8. Consumer perception problems: This situation would be resolved by educating consumers about the safety of the use of phages, and also through the use of new molecules derived from phages, such as endolysins or purified lysozyme (Borysowski and Weber-Dabrowska, 2008). 2.5.

## Efforts Made in Ethiopia Regarding Bacteriophages Therapy

Generally, there is limited study on phage isolation and their therapeutic potential, particularly, against multi-drug resistant pathogen in Ethiopia. However, bacteriophage having ability of lysing MDR Escherichia coli was isolated and demonstrated by Getachew and his co-worker. Moreover, very few review articles on phages, which are done by some universities from Ethiopia, were existed in scientific journals. Efforts made in Ethiopia are specifically paying attention on isolation of bacteriophages from natural sources (Nureye et al., 2018)

### 3. CONCLUSION AND RECOMMENDATION

In an era of increasing emergence of multi-drug resistant pathogens and a scarcity of new antimicrobials, there is an urgent need to find antimicrobial alternatives. Phage-based solutions presented here in have potential as therapeutic or prophylactic tools for application in human animal. Bacteriophages have several characteristics that make them potentially attractive therapeutic agents. They are highly specific and very effective in lysing targeted pathogenic bacteria, safe and rapidly modifiable to combat the emergence of newly arising bacterial threats. In addition a large number of publications, some of which are reviewed in this review, suggest that phages may be effective therapeutic agents in selected bacterial infection of animal. Bacteriophage therapy may be in future effective therapeutic tool for management of bacterial infection. Well-organized research and more evidence from highquality clinical trials will be needed to increase confidence in the breadth of their effectiveness.

Depending on the above fact the following points are recommended:

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☐ Well organized research and substantial academic efforts should be made to realize complete phage therapy utilization in veterinary medicine
☐ Facilitating and organizing workshops as well as training to provide information concerning the possible use of bacteriophages should be made
☐ Clear legal regulations must be established to define limitations and the safe use of phage therapy
☐ Lastly, bacteriophage therapy should be enhanced and application should be adopted in developing countries

like Ethiopia

#### **REFERENCES**

- 1. Aarestrup, F.M., Bager, F., Jensen, N.E. (1998): Surveillance of antimicrobial resistance in bacteria isolated from food animals to antimicrobial growth promoters and related therapeutic agents in Denmark. *Apmis.* **106**: 606-22.
- 2. Abedon, S.T. (2006): Phage ecology. In: Calender, R. (Ed.). The Bacteriophages. New York: Oxford University Press, pp. 37-46.
- 3. Abhilash, M., Viday, A.G. and Jagadevi, T. (2009): Bacteriophage therapy: a war against antibiotic resistance bacteria. *International Journal of Complementary and Alternative Medicine*. 7: 191-221.
- Ackermann, H.W. (2005): Bacteriophage classification. In: Kutter, E. and Sulakvelidze, A. (Eds.). Bacteriophages: Biology and Applications. Florida: CRC Press, pp. 67 89.
- 5. Albino, L.A., Rostagno, M.H., Hungaro, H.M., Mendonca, R.S. (2014): Isolation, characterization, and application of bacteriophages for Salmonella spp. biocontrol in pigs. *Foodborne Pathogen and Diseases*. 11: 602-9.
- 6. Andreatti Filho, R.L., Higgins, J.P. and Higgins, S.E. (2007): Ability of bacteriophages isolated from different sources to reduce Salmonella enterica serovar Enteritidis in vitro and in vivo. *Poultry Science*, **86**:1904–9.
- 7. Atterbury, R.J., Dillon, E., Swift, C., Connerton, P.L., Frost, J.A., Dodd, C.E., Rees, C.E. and Connerton, I.F. (2005): Correlation of Campylobacter bacteriophage with reduced presence of hosts in broiler chicken ceca. *Applied and Environmental Microbiology*. 71: 4885–4887.
- 8. Barrow, P., Lovell, M., Berchieri, A. (1998): Use of lytic bacteriophage for control of experimental Escherichia coli septicemia and meningitis in chickens and calves. *Clinical and Diagnosis Laboratory Immunology*. **5**:294-98
- 9. Bikard, D. and Marraffini, L.A. (2011): Innate and adaptive immunity in bacteria: mechanisms of programmed genetic variation to fight bacteriophages. *Current Opinion in Immunology*. **24**: 5-20.
- 10. Boerlin, P. (2010): Implications of antimicrobial agents as therapeutics and growth promoters in food animal production. ASM Press, Washington, DC, United States, Pp. 1-9.
- 11. Bogovazova, G. G., Voroshilova, N. N., Bondarenko, V. M., Gorbatkova, G. A., Afanas'eva, E. V., Kazakova, T. B. (1992): Immunobiological properties and therapeutic effectiveness of preparations from Klebsiella

- bacteriophages. *Journal of Microbiology, Immunology and Infection.* **3**: 30-33.
- 12. Borie, C., Albala, I., Sanchez, P. (2008): Bacteriophage treatment reduces Salmonella colonization of infected chickens. *Avian Diseases*. **52**: 64-67.
- 13. Borie, C., Sanchez, M., Navarro, C. (2009): Aerosol spray treatment with bacteriophages and competitive exclusion reduces Salmonella enteritidis infection in chickens. *Avian Diseases*. **53**:250–54
- 14. Boriea, C., Robesonb, J. and Galarcea, K. (2014): Lytic Bacteriophage In Veterinary Medicine: a therapeutic option against bacterial pathogen. *Arch med vet.* **46**: 167-179
- 15. Borysowski, J. and Weber-Dabrowska, B. (2008): Bacteriophage endolysins as a novel class of antibacterial agents. *Journal of Experimental Biology and Medicine*. **231**: 366-377.
- 16. Bren, L. (2007): Bacteria-eating virus approved as food additive. FDA Consum. **41**: 20-22
- 17. Brüssow, H. (2005): Phage therapy: the Escherichia coli experience. *Journal of Microbiology*, **151**(7): 2133-2140.
- 18. Brüssow, H. and Kutter, E. (2005): Phage ecology. In: Kutter, E. and Sulakvelidze, A. (Eds.). Bacteriophages: Biology and Application. Florida: CRC Press, pp. 129 –163.
- 19. Buckling, A. and Brockhurst, M. (2012): Bacteriavirus coevolution. *Advanced in Experimental Medicine and Biology*, **751**: 347-370
- 20. Callaway, T.R., Edrington, T.S., Brabban, A.D., Anderson, R.C., Rossman, M.L., Engler, M.J., Carr, M.A., Genovese, K.J., Keen, J.E., Looper, M.L., Kutter, E.M. and Nisbet, D.J. (2008): Bacteriophage isolated from feedlot cattle can reduce Escherichia coli O157:H7 populations in gastrointestinal tracts. Foodborne ruminant Pathogens and Disease 5: 183-191.
- 21. Carlton, R.M. (1999): Phage therapy: past history and future prospects. *Archivum Immunologiae et Therapiae Experimentalis*. **47**: 267-274.
- 22. Carlton, R.M., Noordman, W.H., Biswas, B., De Meester, E.D., Loessner, M.J. (2005): Bacteriophage P100 for control of Listeria monocytogenes in foods: genome sequence, bioinformatics analyses, oral toxicity study, and application. *Regular Toxicology and Pharmacology.* **43**: 301-312.
- 23. CDC, (Centers for Disease Control and Prevention) (2012): Preventing emerging infectious diseases: a strategy for the 21st century overview of the

- updated CDC plan. Atlanta, Georgia, USA. Pp. 25-31.
- 24. Ceyssens, P.J. and Lavigne R. (2010): Introduction to bacteriophage biology and diversity. In: Sabour, P. and Griffiths, M. (Eds.). Bacteriophages in the control of food and waterborne pathogens. ASM Press, Washington, DC, USA. Pp. 11-29.
- 25. Cha, S.B., Yoo, A.N., Lee, W.J. (2012): Effect of bacteriophage in enterotoxigenic Escherichia coli (ETEC) infected pigs. *Journal of Veterinary Medical Science*. **74**: 1037-39.
- 26. Dabrowska, K., Switala-Jelen, K., Opolski, A., Weber-Dabrowska, B., Gorski, B. (2005): Bacteriophage penetration in vertebrates. *Journal of Applied Microbiology.* **98**: 7-13.
- 27. Dewangan, G., Kashyap, P.K. and Giri, D.K. (2017): Bacteriophage therapy and their application. *Journal of Cell and Tissue Research*. 17: 6165-61619
- 28. Duckworth, D. H. (1976): "Who discovered bacteriophage?". *Bacteriological Reviews*, **40**(4): 793-802
- 29. EFSA, (European Food Safety Authority) (2018): The European Union summary report on antimicrobial resistance in zoonotic and indicator bacteria from humans, animals and food in 2017. *Journal of European Food Safety Authority*. **17**(2).
- 30. Fauconnier, A. (2017): Regulating phage therapy: The biological master file concept could help to overcome the regulatory challenge of personalized medicines. *EMBO Report.* **18**: 198-200.
- 31. Fiorentin, L., Vieiry, N.D., Barioni, W. Jr. (2005): Oral treatment with bacteriophages reduces the concentration of Salmonella enteritidis PT4 in caecal contents of broilers. *Avian Pathology.* **34**: 25-63.
- 32. Freitag, T., Squires, R.S. and Schmid, J. 2008. Naturally occurring bacteriophages lyse a large proportion of canine and feline uropathogenic Escherichia coli isolates in vitro. *Res Vet Sci.* 85: 1-7.
- 33. Gannon, V.P., Graham, T.A., King, R., Michel, P., Read, S., Ziebell, K. and Johnson, R.P. (2002): Escherichia coli O157:H7 infection in cows and calves in a beef cattle herd in Alberta, Canada. *Epidemiology and Infection*. **129**: 163-172.
- 34. Gill, J.J., Pacan, J.C., Carson, M.E. (2006): Efficacy and pharmacokinetics of bacteriophage therapy in treatment of subclinical Staphylococcus aureus mastitis in lactating dairy cattle. *Antimicrobial Agents and Chemotherapy.* **50**:2912-18

- 35. Gorski, A. and Weber-Dabrowska, B. (2004): The potential role of endogenous bacteriophages in controlling invading pathogens. *Cellular and Molecular Life Science*. **62**: 511-519
- 36. Hanlon, G.W. (2007): Bacteriophages: an appraisal of their role in the treatment of bacterial infections. *International Journal of Antimicrobial Agents*. **30**:118-28
- 37. Hao, H., Cheng, G., Iqbal, Z. (2014): Benefits and risks of antimicrobial use in food-producing animals. Front Microbiology. **5**:288.
- 38. Harper, D.R. (2006): Biological control by microorganisms. The Encyclopedia of Life Sciences. John Wiley & Sons Chichester 1-10.
- 39. Hawkins, C., Harper, D., Burch, D., Anggard, E., Soothil, J. (2010): Topical treatment of Pseudomonas aeruginosa otitis of dogs with a bacteriophage mixture: A before/after clinical trial. *Vet Microbiology*. **146**: 309-313.
- 40. Higgins, S., Higgins, J.P., Bielke, I., Hargis, B. (2007): Selection and application of bacteriophages for treating Salmonella Enteritidis infections in poultry. *International Journal of Poultry Science*. **6**: 163-168.
- 41. Huff, W., Huff. G., Rath .N. (2003): Evaluation of aerosol spray and intramuscular injection of bacteriophage to treat an Escherichia coli respiratory infection. *Poultry Science.* **82**:1108-12.
- 42. Huff, W.E., Huff G.R., Rath, N.C. (2004): Therapeutic efficacy of bacteriophage and Baytril (enrofloxacin) individually and in combination to treat colibacillosis in broilers. *Poultry Science*. **83**:1944-47.
- 43. Huff, W.E., Huff, G.R., Rath, N.C. (2002): Prevention of Escherichia coli respiratory infection in broiler chickens with bacteriophage (SPR02). *Poultry Science.* **81**: 437-41.
- 44. ICTV, (International Committee on Taxonomy of Virus) (2011): Virus Taxonomy: Release virus (current). http://ictvonline.org/virus Taxonomy. Ackermann, H. W. and Prangishvili, D. (2012): Prokaryote viruses studied by electron microscopy. *Archives of Virology*. **157**(10): 1843–1849.
- 45. Jamalludeen, N., Johnson, R.P., Shewen, P.E., Gyles, C.L. (2009): Evaluation of bacteriophages for prevention and treatment of diarrhea due to experimental enterotoxigenic Escherichia coli O149 infection of pigs. *Veterinary Microbiology*. **136**: 135-41
- 46. Keen, E.C. (2012): Felix d'Herelle and Our Microbial Future. *Future Microbiology*. **7**(12): 1337–1339.

- 47. Korotyayev, A. and Babichev, A. (2002): Medical microbiology, immunology and virology, 5<sup>th</sup>ed. London: Oxford University Press.
- 48. Kutateladze, M. (2015): Experience of the Eliava Institute in bacteriophage therapy. *Journal of Virology*. **30**: 80-81.
- 49. Kutter, E. and Sulakvelidze, A. (2005): Bacteriophages: biology and applications. CRC Press, Boca Raton, Fla, USA
- 50. Lee, N. and Harris, D. (2001): The effect of bacteriophage treatment to reduce the rapid dissemination of Salmonella typhimurium in pigs. Swine Res Report. Pp 50.
- 51. Leitenberg, M. (2001): Biological weapons in the twentieth century: a review and analysis. *Journal of Critical Review of Microbiology*. **27**: 267-320.
- 52. Lim, T.H., Kim, M.S., Lee, D.H. (2012): Use of bacteriophage for biological control of Salmonella Enteritidis infection in chicken. *Research in Veterinary Science.* **93**:1173-78
- 53. Lima, D., Furian, T., Pillati, R. (2016): Establishment of a pathogenicity index in Salmonella Enteritidis and Salmonella Typhimurium strains inoculated in one-day-old broiler chicks. *Journal of Veterinary Medicine and Animal Health.* 68: 257-64
- 54. Loc-Carrillo, C. and Abedon, S.T. (2001): Pros and cons of phage therapy. *Bacteriophage*. 1: 111-114.
- 55. Luria, S.E., Delbrück, M., Anderson, T.F. (1993): Electron Microscope Studies of Bacterial Viruses. *International Journal of Bacteriology*. **46**: 57-77.
- 56. Machado, V.S., Bicalho, M.S., Pereira, R.V., Caixeta, L.S., Bittar, J.H., Oikonomou, J.G., Gilbert, O.R., Bicalho, R.C. 2012. The effect of intrauterine administration of mannose or bacteriophage on uterine health and fertility of dairy cows with special focus on Escherichia coli and Arcanobacterium pyogenes. *Journal of Dairy Science*. 95: 3100-3109.
- 57. Mellon, M., Benbrock, C. and Benbroc, K. (2001): Hogging it: estimates of antimicrobial abuse in livestock. Cambridge (MA): Union of Concerned Scientists.
- Merril, C. R. (2008): Interaction of Bacteriophages with Animals. In: Bacteriophage Ecology, Abedon,
  S. T. (Ed.). Cambridge University Press: Cambridge, UK, pp. 332-352.
- 59. Monk, A.B., Rees, C.D., Barrow, P., Hagens, S. and Harper, D.R. (2010): Bacteriophage applications: where are we now? *Letters in Applied Microbiology*. **51**: 363
- 60. Nakai, T. (2010): Application of bacteriophages for control of intectious diseases in aquaculture. In:

- Sabour, P.M. and Griffiths, M.W. (Eds.). Bacteriophage in the Control of Food and Waterborne Pathogens. ASM Press, Washington DC, USA, Pp 257-272
- 61. Newell, D.G., Koopmans, M., Verhoef, L., Duizer, E., Aidara-Kane, A., Sprong, H., Opsteegh, M., Langelaar, M., Threfall, J., Scheutz, F., Van Der Giessen, J. and Kruse, H. (2010): Food-borne diseases: the challenges of 20 years ago still persist while new ones continue to emerge. *International Journal of Food Microbiology*. 139: 3-15
- 62. Niu, Y.D., Xu, Y., McAllister, T.A., Rozema, E.A., Stephens, T.P., Bach, S.J., Johnson, R.P. and Stanford, K, (2008): Comparison of fecal versus rectoanal mucosal swab sampling for detecting Escherichia coli O157:H7 in experimentally inoculated cattle used in assessing bacteriophage as a mitigation strategy. *Journal of Food Protection*. 71: 691-698
- 63. Nureye, D., Mohammed, S. and Assefa, S. (2018): Bacteriophages and its applications: general aspects, and a new insight in Ethiopia. *Journal of Drug Delivery and Therapeutics*. **8**(6): 278-284
- 64. O'Flaherty, S., Coffey, A., Meaney, W.J. (2005). Inhibition of bacteriophage K proliferation on Staphylococcus aureus in raw bovine milk. *Letters in Applied Microbiology*. **41**:274–79
- 65. Oliveira, A., Sereno, R., Azeredo, J. (2010): In vivo efficiency evaluation of a phage cocktail in controlling severe colibacillosis in confined conditions and experimental poultry houses. *Veterinary Microbiology*. 146: 303-8.
- 66. Page, S. and Gautier, P. (2012): Use of antimicrobial agents in livestock. *Rev Sci Tech.* **31**: 145 88.
- 67. Parisien, A., Allain, B., Zhang, J., Mandeville, R. and Lan, Q. (2008): Novel alternatives to antibiotics: bacteriophages, bacterial cell wall hydrolases, and antimicrobial peptides. *Journal of Applied Microbiology*. **104**(1):1–13.
- 68. Payne, R.J. and Jansen, V.A. (2001): Understanding bacteriophage therapy as a density-dependent kinetic process. *Journal of Theoretical Biology*. **208**: 37-48.
- 69. Pereira, C., Silva, Y.J., Santos, A.L., Cunka, A., Gomes, M.G., Almeida, A. (2011): Bacteriophages with potential for inactivation of fish pathogenic bacteria: survival, host specifies and effect on bacterial community structure. *Mar Drugs*. **9**: 2236-2255.
- 70. Pimenov, N. and Danilevskaya, N. (2006): The antibiotic resistance of Salmonella, isolated from pigeons. *The Veterinary Journal*. **9**: 25-45.

- 71. Rangel, J.M., Sparling, P.H., Crowe, C., Griffin, P.M. and Swerdlow, D.L. (2005): Epidemiology of Escherichia coli O157:H7 outbreaks, United States, 1982–2002. *Emerging Infectious Diseases* 11: 603-609
- 72. Rozema, E.A., Stephens, T.P., Bach, S.J Okine, E.K., Johnson, R.P., Stanford, K., Mcallister, T.A. (2009): Oral and rectal administration of bacteriophages for control of Escherichia coli O157:H7 in feedlot cattle. *Journal of Food Protect*. 72: 241-250.
- 73. Ryan, E.M., Gorman, S.P., Donnelly, R.F. and Gilmore, B.F. (2001): Recent advances in bacteriophage therapy: how delivery routes, formulation, concentration and timing influence the success of phage therapy. *Journal of Pharmacy and Pharmacology*. **63**: 1253-1264
- 74. Saez, A.C., Zhang, J., Rostagno, M.H., Ebner, P.D. (2011): Direct feeding of microencapsulated bacteriophages to reduce Salmonella colonization in pigs. *Foodborne Pathogen and Diseases*. **8**:1269-74
- 75. Santos, T.A., Ledbetter, E.C., Caixeta, L.S., Bicalho, M.L., Bicalho, R.C. (2011): Isolation and characterization of two bacteriophages with strong in vitro antimicrobial activity against Pseudomonas aeruginosa isolated from dogs with ocular infections. *American Journal of Veterinary Research.* 72: 1079-1086.
- 76. Sheng, H., Knecht, H.J., Kudva, I.T. and Hovde, C.J. (2006): Application of bacteriophages to control intestinal Escherichia coli O157:H7 levels in ruminants. *Applied and Environmental Microbiology* 72: 5359–5366.
- 77. Sklar, I.B. and Joerger RD. (2001): Attempts to utilize bacteriophage to combat Salmonella enterica serovar enteritidis infection in chickens. *Journal of Food Safety.* **21**:15-29.
- 78. Skurnik, M. and Strauch, E. (2006): Phage therapy: facts and fiction. *Journal Medical Microbiology*. **296**: 5-14
- 79. Smith, H. and Huggins, M. (1983). Effectiveness of phages in treating experimental Escherichia coli diarrhea in calves, piglets and lambs. *Journal of General Microbiology*. **129**:2659-75.
- 80. Sulakvelidze, A. and Kutter, E. (2005): Bacteriophage therapy in humans. In: Kutter, E. and Sulakvelidze, A. (Ed.). Bacteriophages: Biology and Applications. CRC Press, Boca Raton, USA, Pp 381-436.
- 81. Sulakvelidze, A. and Kutter, E. (2012) Bacteriophages: Biology and Applications, 1st ed.

- 82. Sulakvelidze, A. and Morris, J. G. (2001): Bacteriophages as therapeutic agents. *Journal of Annals of Medicine*. **33**: 507-509.
- 83. Sulakvelidze, A., Alavidze, Z., Morris, J. (2001): Bacteriophage therapy. *Antimicrobial Agents and Chemotherapy.* **45**: 649-659
- 84. Summers, W. C. (2001): Bacteriophage therapy. *Annual Review of Microbiology*. **55**: 437–451.
- 85. Summers, W.C. (1999): Felix d'Herelle and the origins of molecular biology. Yale University Press, New Haven, Conn.
- 86. Toro, H., Price, S., McKee, A. (2005): Use of bacteriophages in combination with competitive exclusion to reduce Salmonella from infected chickens. *Avian Diseases*. **49**:118-24.
- 87. Uchiyama, J., Matsuzaki, S. and Rashel, M. (2005): Bacteriophage therapy: a revitalized therapy against bacterial infectious diseases. *Journal of Infection and Chemotherapy.* **11**(5): 211–219.
- 88. Verbeken, G., De Vos, D., Vaneechoutte, M., Merabishvil, M., Zizi, M. and Prinay, J-P. (2007): European regulatory conundrum of phage therapy. *Future Microbiology*. **2**: 485-491
- 89. Wagenaar, J.A., Van Bergen, M.A., Mueller, M.A., Wassenaar, T.M., Carlton, R.M. (2005): Phage therapy reduces Campylobacter jejuni colonization in broilers. *Veterinary Microbiology*. **109**: 2
- 90. Waldor, M. K., Friedman, D., Adhya, S. (2005): Phages: Their Role in Bacterial Pathogenesis and Biotechnology; ASM Press: Washington, DC.

- 91. Wall, S.K., Zhang, J., Rostagno, M.H., Ebner, P.D. (2010): Phage therapy to reduce preprocessing Salmonella infections in market-weight swine. *Applied and Environmental Microbiology*. **76**:48-53.
- 92. Waxman, D.J. and Strominger, J.L. (1983): Penicillin-binding proteins and the mechanism of action of beta-lactam antibiotics. *Annular Review of Biochemistry*. **52**: 825-869.
- 93. Wernicki, A., Nowaczek, A., Urban-Chmiel, R. (2017): Bacteriophage therapy to combat bacterial infections in poultry. *Journal of Virology*. **14**(1): 75-283
- 94. WHO, (World Health Organization) (2000): Overcoming antimicrobial resistance, World Health Organization (Report on Infectious Diseases), Geneva, Switzerland. Pp. 11-29.
- 95. WMA Declaration of Helsinki. (2008): Ethical Principles for Medical Research Involving Human Subjects. Adopted by then18th WMA General Assembly, Helsinki, Finland and amended by the: 59th WMA General Assembly, Seoul, Korea, paragraph 32.
- 96. Yoshida, H., Nakamura, M., Bogaki, M., Ito, H., Kojima, T. (1993) Mechanism of action of quinolones against Escherichia coli DNA gyrase. *Antimicrobial Agents and Chemotherapy.* **37**: 839-845.

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